

Prolonged SARS-CoV-2 shedding in saliva; implications for late-stage diagnosis and infectious duration

Abby Chopoorian^{1,2}, Padmapriya Banada¹, Robert Reiss¹, David Elson^{1,2}, Claire Park^{1,2}, Sukalyani Banik¹, Naranjargal Daivaa¹, Soyi Sarkar¹, Emily Hennig³, Austin Togba³, Aanchal Wats³, Abraham Wei¹, Laura Palo², Mitchell Hirsch², Carter Campbell², Pooja Saiganesh², David Alland¹, Yingda L. Xie¹

¹ Public Health Research Institute and Division of Infectious Diseases, Department of Medicine, Rutgers New Jersey Medical School, Newark, NJ, USA

² School of Medicine, Rutgers New Jersey Medical School, Newark, NJ, USA

³ Rutgers School of Public Health, Piscataway, NJ, USA

Background

- PCR testing of saliva has been shown to have comparable (1-3) and even higher sensitivity (4) and stability (5) than PCR testing of nasopharyngeal (NP) at identifying COVID-19
- Dynamics of early viral shedding from saliva can differ from nasal specimens (6), guiding preferences for testing strategies at different stages of infection
- Viral shedding in saliva may be more stable than nasal swabs during the late post-symptomatic period
- We conducted a sub-study within a longitudinal observational study to compare the longevity of viral shedding from the mouth versus nasal swabs

Methods

- **Study population:** patients with moderate to severe COVID-19 at University Hospital from June 2020 – August 2021
- **Data collection:** Saliva, anterior nasal swabs, and nasopharyngeal swabs collected and tested with Cepheid Xpert Xpress SARS-CoV-2 RT-PCR assay to determine cycle threshold (Ct)
- **Inclusion Criteria:** ≥18 years, COVID-19 PCR positive
- **Exclusion criteria:** prisoners, unknown time from symptom onset, unable to provide both saliva and nasal specimens
- **Statistical Analysis:**
 - Median Ct bias-corrected and accelerated 95% confidence intervals via bootstrapping and compared using Wilcoxon signed-rank
 - Patient characteristic differences via ANOVA for continuous and chi-square test for categorical measures

Results

Table 1. Patients' characteristics

Symptom duration prior to baseline collection Mean (range)	7.0 days (-1 – 21)	8.6 days (2 – 21)
Days between in-hospital NP swab PCR and baseline collection: mean (range)	1.8 days (-4 – 7)	1.7 days (0 – 6)
Number of follow-up time-points per participant: mean (range)	0.5 timepoints (0 - 7)	1.8 timepoints (1 - 7)
# Male (%) # Female (%)	51 (53%) 45 (47%)	18 (64%) 10 (36%)
Ethnicity (%)		
Hispanic	53 (55%)	17 (61%)
Black	36 (38%)	7 (25%)
White	4 (4%)	2 (7%)
More than one race/Other	3 (3%)	2 (7%)
Comorbidities		
Hypertension	51 (53%)	14 (50%)
Obesity	48 (50%)	12 (46%)
Diabetes Mellitus	33 (34%)	10 (36%)
Lung Disease (eg, COPD)	12 (13%)	5 (18%)
Chronic Heart Disease	10 (10%)	4 (14%)
Other	41 (43%)	9 (32%)
No chronic disease	18 (19%)	3 (11%)

Figure 2. Survival Curve

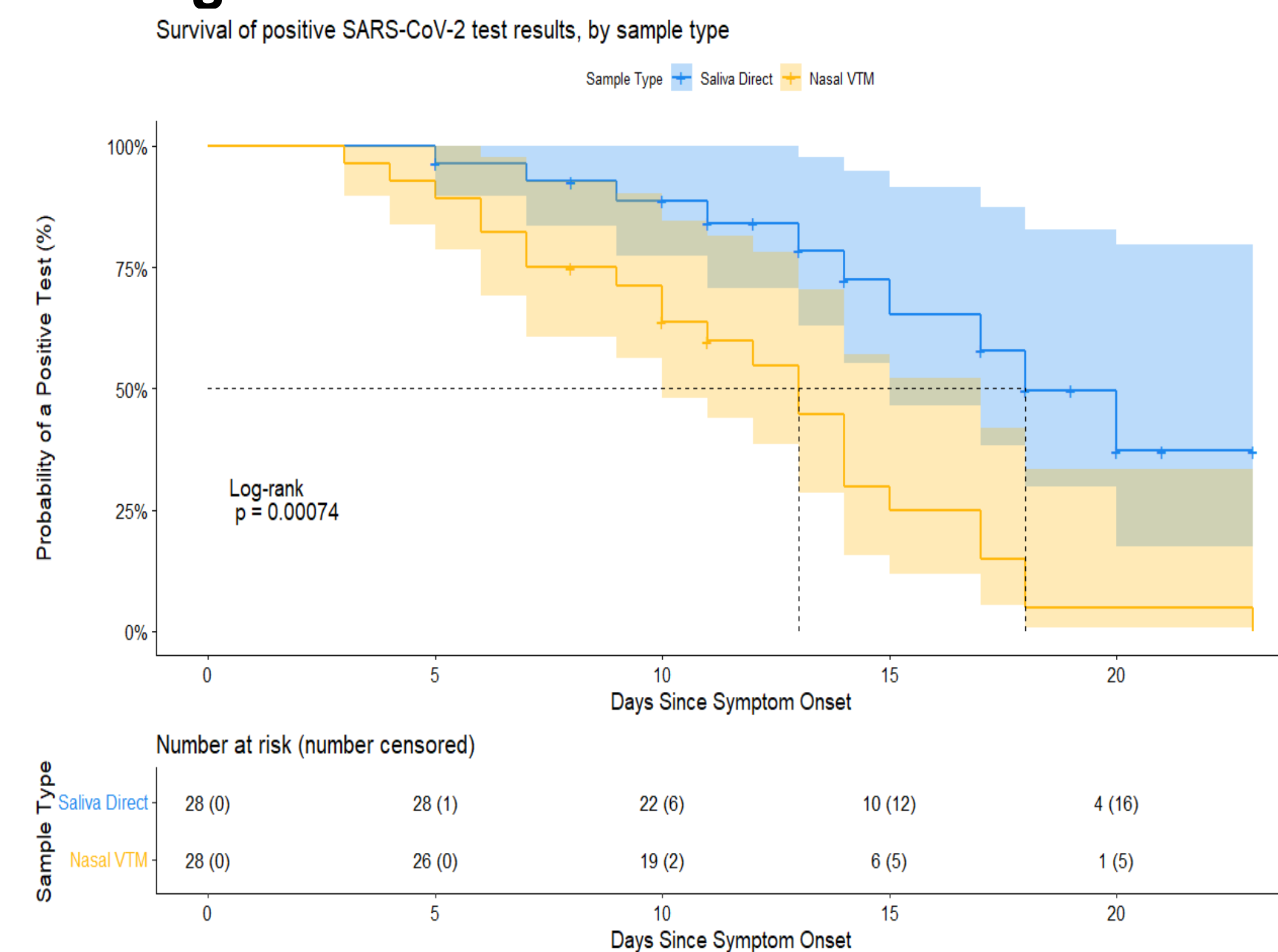
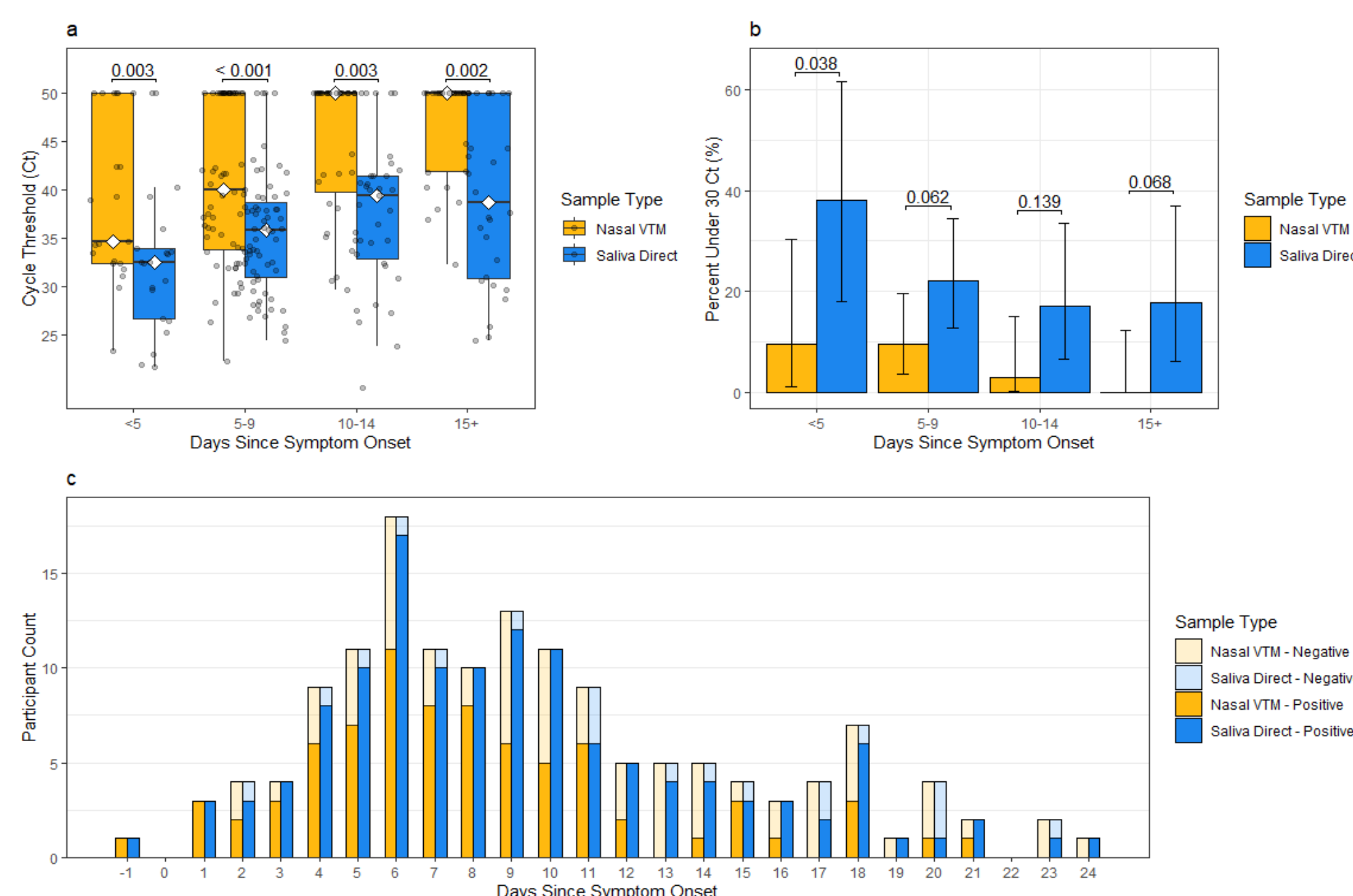


Figure 1. Nasal VTM vs Saliva



- Saliva detected significantly more cases of SARS-CoV-2 **beyond 5 days** of symptom onset (n = 115, 86.1% saliva vs 48.7% nasal, p-value < 0.001), **beyond 9 days** (n = 63, 79.4% saliva vs 36.5% nasal, p-value < 0.001) and **beyond 14 days** of (n = 28, 71.4% saliva vs 32.1% nasal, p-value = 0.003)

Conclusion

- **Study limitations:**
 - Conducted prior to the emergence of the Omicron variant (dynamics of viral shedding may vary with variants of concern)
 - Small sample size
 - Limited longitudinal data
- **Key Findings:**
 - SARS-CoV-2 RNA shedding persists longer and in higher abundance in saliva than in nasal swabs, even beyond 14 days
 - PCR testing of saliva may be more sensitive than nasal swabs in diagnosing COVID in patients who present during a later disease stage
 - Public health implications include potential prolonged oral infectious period in guidance for masking and follow-up saliva-based testing for known COVID-19
- **Funding source:** Rutgers Center for COVID-19 Research and Pandemic Preparedness (CCRP2)

References

1. Wylie AL, Fournier J, Casanovas-Massana A, Campbell M, Tokuyama M, Vijayakumar P, et al. Saliva is more sensitive for SARS-CoV-2 detection in COVID-19 patients than nasopharyngeal swabs [Internet]. Infectious Diseases (except HIV/AIDS); 2020 Apr [cited 2022 Mar 17]. Available from: <http://medrxiv.org/lookup/doi/10.1101/2020.04.16.20067835>
2. Banada P, Elson D, Daivaa N, Park C, Desind S, Montalvan I, et al. Evaluation of sample collection and transport strategies to enhance yield, accessibility, and biosafety of COVID-19 RT-PCR testing [Internet]. Infectious Diseases (except HIV/AIDS); 2021 Mar [cited 2022 Mar 17]. Available from: <http://medrxiv.org/lookup/doi/10.1101/2021.03.03.21251172>
3. Kojima N, Turner F, Slepnev V, Bacelar A, Deming L, Kodeboyina S, et al. Self-Collected Oral Fluid and Nasal Swabs Demonstrate Comparable Sensitivity to Clinician Collected Nasopharyngeal Swabs for Covid-19 Detection [Internet]. Infectious Diseases (except HIV/AIDS); 2020 Apr [cited 2022 Mar 17]. Available from: <http://medrxiv.org/lookup/doi/10.1101/2020.04.11.20062372>
4. Teo AKJ, Choudhury Y, Tan IB, Cher CY, Chew SH, Wan ZY, et al. Saliva is more sensitive than nasopharyngeal or nasal swabs for diagnosis of asymptomatic and mild COVID-19 infection. Sci Rep. 2021 Dec;11(1):3134.
5. Wylie AL, Fournier J, Casanovas-Massana A, Campbell M, Tokuyama M, Vijayakumar P, et al. Saliva or Nasopharyngeal Swab Specimens for Detection of SARS-CoV-2. N Engl J Med. 2020 Sep 24;383(13):1283–6.
6. Lai J, German J, Hong F, Tai SHS, McPhaul KM, Milton DK, et al. Comparison of Saliva and Mid-Turbinate Swabs for Detection of COVID-19 [Internet]. Infectious Diseases (except HIV/AIDS); 2021 Dec [cited 2022 Mar 17]. Available from: <http://medrxiv.org/lookup/doi/10.1101/2021.12.01.21267147>